

MAPK Signaling Is Critical to Estradiol Protection of CA1 Neurons in Global Ischemia

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The importance of hormone therapy in affording protection against the sequelae of global ischemia in postmenopausal women remains controversial. Global ischemia arising during cardiac arrest or cardiac surgery causes highly selective, delayed death of hippocampal CA1 neurons. Exogenous estradiol ameliorates global ischemia-induced neuronal death and cognitive impairment in male and female rodents. However, the molecular mechanisms by which estrogens intervene in global ischemia-induced apoptotic cell death are unclear. Here we show that estradiol acts via the classical estrogen receptors, the IGF-I receptor, and the ERK/MAPK signaling cascade to protect CA1 neurons in ovariectomized female rats and gerbils. We demonstrate that global ischemia promotes early dephosphorylation and inactivation of ERK1 and the transcription factor cAMP-response element binding protein

(CREB), subsequent down-regulation of the antiapoptotic protein Bcl-2, a known gene target of estradiol and CREB, and activation of caspase-3. Estradiol treatment increases basal phosphorylation of both ERK1 and ERK2 in hippocampal CA1 and prevents ischemia-induced dephosphorylation and inactivation of ERK1 and CREB, down-regulation of the caspase death cascade. Whereas ERK/MAPK signaling is critical to CREB activation and neuronal survival, the impact of estradiol on Bcl-2 levels is ERK independent. These findings support a model whereby estradiol acts via the classical estrogen receptors and IGF-I receptors, which converge on activation of ERK/MAPK signaling and CREB to promote neuronal survival in the face of global ischemia. (*Endocrinology* 148: 1131–1143, 2007)

GLOBAL BRAIN ISCHEMIA, arising during cardiac arrest, open heart surgery, profuse bleeding, or experimental induction in animals via bilateral carotid artery occlusion, causes selective, delayed neuronal death and delayed neurological deficits (reviewed in Ref. 1). Pyramidal neurons in the hippocampal CA1 are particularly vulnerable, whereas interneurons in this cell layer and pyramidal neurons in other hippocampal subfields survive. Histological evidence of CA1 pyramidal neuron degeneration is not observed until 2–3 d after global ischemia in rats or 3–4 d in gerbils (2–5). Although the mechanisms underlying ischemia-induced death are as yet unclear, the substantial delay between insult and onset of death provides the opportunity to examine molecular events that destine these neurons to die.

Estradiol-17 β , the primary estrogen secreted by the ovaries, acts on neurons to increase spine density and synapse number (6–8) and *N*-methyl-D-aspartate receptor NR1 subunit expression (9) and potentiates kainate-elicited currents in CA1 pyramidal neurons (10, 11). Moreover, estrogens afford neuroprotection in experimental models of global and focal ischemia (12–15) and ameliorate the cognitive deficits associated with ischemic cell death (16, 17). Although the cellular sites mediating these actions are unclear, estrogen receptors- α and - β are expressed in the hippocampus in

which they could subserve the neuroprotective actions of estradiol (18).

Cross talk between estradiol and trophic factors such as IGF-I is implicated in the cellular actions of estradiol. Estradiol and IGF-I act synergistically in neurons to regulate synaptic remodeling, neuronal differentiation and neuronal survival (19). IGF-I receptors are critical to estradiol protection of hilar neurons from seizure-induced injury (20). In the brain, estradiol and IGF-I activate ERK/MAPK (19), a well-characterized intracellular signaling cascade implicated in neuronal plasticity and survival (13, 21, 22). Upon stimulation with estradiol, estradiol receptor- α and IGF-I receptor form a macromolecular signaling complex, which recruits and activates downstream kinases including MAPK (23–25). ERK/MAPK signaling culminates in phosphorylation and activation of nuclear transcription factors such as cAMP-response element binding protein (CREB), which regulate target genes important to neuronal survival and protection. CREB targets implicated in neuronal survival include the antiapoptotic protein Bcl-2 and brain-derived neurotrophic factor (BDNF) (21).

The present study sought to identify molecular targets, especially intracellular signaling cascades, which mediate estradiol neuroprotection in global ischemia. We show that estradiol acts via classical estrogen receptors, IGF-I receptors, and ERK/MAPK signaling to promote neuronal survival after transient global ischemia. Global ischemia promotes dephosphorylation and inactivation of ERK1 and its target, the transcription factor CREB, followed by down-regulation of Bcl-2 and activation of the caspase death cascade. Estradiol markedly attenuates ischemia-induced dephosphorylation and inactivation of ERK1 and CREB, down-regulation of Bcl-2 and activation of caspase-3. Pharmacological blockade

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Abbreviations: BDNF, Brain-derived neurotrophic factor; CREB, cAMP-response element binding protein; DMSO, dimethylsulfoxide; FAM, carboxyfluorescein; FMK, fluoromethyl ketone; GPR, G protein-coupled receptor; icv, intracerebroventricular; MEK, MAPK kinase.

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of ERK reverses the effects of estradiol on both ERK and CREB phosphorylation but not on Bcl-2. Thus, estradiol and IGF-I coordinately activate ERK/MAPK signaling and its target CREB, thereby promoting neuronal survival in the face of global ischemia.

Materials and Methods

Animals

Age-matched adult female Mongolian gerbils weighing 60–80 g (Tumblebrook Farms, Wilmington, MA) and female Sprague Dawley rats weighing 100–150 g (Charles River, Wilmington, DE) at the time of ischemic insult were maintained in a temperature- and light-controlled environment with a 14-h light, 10-h dark cycle and were treated in accordance with the principles and procedures of the National Institutes of Health (NIH) Guidelines for the Care and Use of Laboratory Animals. Protocols were approved by the Institutional Animal Care and Use Committee of the Albert Einstein College of Medicine.

Ovariectomy and estradiol pellet implantation

Female Mongolian gerbils were ovariectomized and female Sprague Dawley rats were ovariectomized under halothane anesthesia (4% for induction, followed by 1% for maintenance). Seven days later, pellets containing estradiol-17 β (gerbils: 0.36 mg/pellet, 60 d sustained release; rats: 0.05 mg/pellet, 21 d sustained release; Innovative Research of America, Inc., Sarasota, FL) or placebo were inserted sc beneath the dorsal surface of the neck. Estradiol pellets were present for 14 d before ischemia and remained in place until the animals were killed at 1 h to 2 d after ischemia for molecular studies or 7 d after ischemia for histological analysis.

Global ischemia

Experiments were performed in rats, with the exception of the estrogen receptor and IGF-I receptor antagonist treatments, which were performed in gerbils. Gerbils offer an advantage, compared with rats, in that they lack posterior communicating arteries, structures that in humans and rats complete the circle of Willis and permit collateral blood flow via the vertebral arteries. Thus, global ischemia can be produced in gerbils by the relatively simple procedure, bilateral occlusion of the carotid arteries. However, many of the available antibodies used in the present study recognize rat (but not gerbil) proteins with high affinity and good signal to noise. Therefore, we carried out most experiments in rats using four-vessel occlusion (see below).

Fourteen days after pellet implantation, gerbils were fasted overnight and anesthetized with halothane (4% for induction, followed by 1% for maintenance) delivered by mask in a mixture of N₂/O₂ (70:30) by means of a Vapomatic anesthetic vaporizer (CWE Inc., Ardmore, PA). Gerbils were subjected to global ischemia by temporary occlusion of both carotid arteries for 5 min with nontraumatic aneurism clips followed by reperfusion as described (26) or to sham operation. Sham-operated animals were subjected to the same anesthesia and surgical procedures as animals subjected to global ischemia, except that the carotid arteries were not occluded. Rats were subjected to global ischemia by four-vessel occlusion as described (27). In brief, 13 d after pellet implantation, rats were fasted overnight and anesthetized with halothane as described above for gerbils. The vertebral arteries were subjected to electrocauterization, the common carotid arteries were exposed and isolated with a 3–0 silk thread, and the wound was sutured. Twenty-four hours later, the animals were anesthetized again, the wound was reopened and both carotid arteries were occluded for 10 min with nontraumatic aneurism clips, followed by reperfusion. Arteries were visually inspected to ensure adequate reflow. Sham-operated rats were subjected to the same anesthesia and surgical procedures as animals subjected to global ischemia (vertebral artery coagulation and carotid artery exposure), except that the carotid arteries were not occluded.

In all cases, anesthesia was discontinued immediately after initiation of carotid artery occlusion. Body temperature was monitored and maintained at 37.5 \pm 0.5 C with a rectal thermistor and heat lamp until recovery from anesthesia. Animals that failed to show complete loss of the righting reflex and dilation of the pupils from 2 min after occlusion

was initiated until the end of occlusion and the rare animals that exhibited obvious behavioral manifestations (abnormal vocalization when handled, generalized convulsions, hypoactivity) or loss of more than 20% body weight by 3–7 d were excluded from the study. After reperfusion, arteries were visually inspected to ensure adequate flow. Sixty gerbils and 175 rats were subjected to global ischemia. There were seven deaths in gerbils and three deaths in rats due to respiratory arrest. Another two gerbils and 18 rats were excluded from the study because they failed to show neurological signs of ischemia (no loss of consciousness or incomplete dilation of the pupils during occlusion). Rats and gerbils that died from respiratory arrest or failed to show neurological signs of ischemia were not concentrated in either the placebo or estradiol-treated groups.

ICI 182,780, JB-1, and PD98059 intracerebroventricular (icv) administration

Halothane-anesthetized animals were injected with the broad-spectrum estrogen receptor antagonist ICI 182,780 (100 μ g; Tocris, Ellisville, MO), the competitive IGF-I receptor antagonist JB-1 (10 μ g; Bachem Inc., Budendorf, Switzerland), or the ERK/MAPK inhibitor PD98059 (3 μ g; Calbiochem, La Jolla, CA) or vehicle (10 μ l of 50% dimethylsulfoxide (DMSO), saline, or 10% DMSO, respectively) by unilateral injection into the right lateral ventricle at a flow rate of 5 μ l/min immediately after global ischemia (14 d after pellet implantation) or sham surgery and again 12 h later. The icv injections of 75% DMSO have no obvious harmful effects (28). Animals were positioned in a Kopf small animal stereotaxic frame with the incisor bar lowered 3.3 \pm 0.4 mm below horizontal zero. A stainless steel cannula (28 gauge) was lowered stereotaxically into the right lateral ventricle to a position defined by the following coordinates: 0.4 mm posterior to bregma, 1.2 mm lateral to bregma, 2.6 mm below the skull surface (gerbils), or 0.92 mm posterior to bregma, 1.2 mm lateral to bregma, 3.6 mm below the skull surface (rats) according to the atlas of Paxinos and Watson (29).

Histological analysis

Neuronal cell loss was assessed by histological examination of toluidine blue-stained brain sections at the level of the dorsal hippocampus from animals killed at 7 d after ischemia (21 d after pellet implantation) or sham operation. Animals were deeply anesthetized with pentobarbital (50 mg/kg, ip), and blood was collected by cardiac puncture for assay of plasma estradiol levels (see below); this was followed by transcardiac perfusion with ice-cold 4% paraformaldehyde in PBS [0.1 M (pH 7.4)]. Brains were removed and immersed in fixative (4 C overnight). Coronal sections (15 μ m) were cut at the level of the dorsal hippocampus (3.3–4.0 mm posterior from bregma) with an electronic cryostat (Thermo Electron Corp., Pittsburgh, PA), and every fourth section was collected and stained with toluidine blue. The number of surviving pyramidal neurons per 250- μ m length of the medial CA1 pyramidal cell layer was counted bilaterally in four sections per animal as described (30) under a light microscope at \times 40 magnification. Cell counts from the right and left hippocampus on each of the four sections were averaged to provide a single value (number of neurons per 250 μ m length) for each animal.

Serum estradiol assay

Tubes containing whole blood were placed on ice (10 min) and centrifuged at 300 \times g for 5 min. Serum was collected and stored (-20 C) until analyzed. Serum hormone levels were measured by fluoroimmunoassay using the DELPHIA estradiol assay (PerkinElmer Life Sciences, Turku, Finland). All assays were performed in duplicate and the mean value reported. The sensitivity of detection is 13 pg/ml. The inter- and intraassay coefficients of variance are 10.1 and 4.1%, respectively.

Antibodies

The following antibodies were used in this study: 1) Bcl-2, an affinity purified polyclonal antibody raised against a peptide mapping within the amino terminus of Bcl-2 of human origin (1:100 for immunolabeling, 1:1000 for Western; Santa Cruz Biotechnology, Inc., Santa Cruz, CA); 2) antiphospho MAPK (p-ERK1/2) mouse monoclonal antibody, which

recognizes ERK1 and ERK2 that are phosphorylated on both a threonine and a tyrosine residue (clone 12D4, 1:5000; Upstate Biotechnology, Inc., Lake Placid, NY); 3) anti-MAPK1/2 (ERK1/2) rabbit polyclonal antibody, which recognizes total ERK1/ERK2 (1:5000; Upstate Biotechnology); 4) anti-ERK5 and phospho-ERK5 (p-Thr218/p-Tyr220) rabbit polyclonal antibodies (ERK5, 1:500; p-ERK5, 1:200; Cell Signaling Technology, Inc., Beverly, MA); 5) antiphospho-CREB (p-CREB) rabbit polyclonal antibody, which recognizes CREB phosphorylated at serine 133 (1:3000; Upstate Biotechnology); 6) anti-CREB rabbit polyclonal antibody, which recognizes total CREB (1:3000; Upstate Biotechnology); 7) anti- β -actin mouse monoclonal antibody, which recognizes an epitope located within the N-terminal domain of the β -isoform of actin (1:20,000; Sigma, St. Louis, MO); and 8) antihistone H3 rabbit polyclonal antibody, which recognizes endogenous histone H3 protein only (1:1000; Cell Signaling Technology). Secondary antibodies for Westerns were horseradish peroxidase-conjugated donkey antirabbit IgG (1:5000; Amersham, Buckinghamshire, UK) for polyclonal antibodies or sheep anti-mouse IgG (1:2500; Amersham) for monoclonal antibodies.

Western blot analysis

For quantification of protein abundance in the hippocampal CA1, whole-cell lysates (ERK1/2, p-ERK1/2), cytosolic (Bcl-2), and nuclear (CREB, p-CREB) fractions isolated from the microdissected hippocampal CA1 subfield of experimental and sham animals were prepared at 1, 3, 12, 24, and 48 h after reperfusion. Proteins were separated by SDS-PAGE and subjected to Western blot analysis as described (27). Protein concentration was determined by BCA protein assay kit (Pierce, Rockford, IL). Aliquots of protein (40–70 μ g) were dissolved in Laemmli sample buffer [0.025 M Tris-HCl, 5% glycerol, 1% sodium dodecyl sulfate, 0.5% PBS, 0.1 M dithiothreitol, 2.5 mM β -mercaptoethanol, 1 mM phenylmethylsulfonyl fluoride, 0.5 mM NaHNO₃ buffer (pH 6.8)], loaded onto 10% polyacrylamide gels, subjected to electrophoresis, and transferred to nitrocellulose membranes for immunolabeling with antibodies to p-ERK1/2, ERK1/2, p-ERK5, ERK5, p-CREB, CREB, or Bcl-2. After reaction, membranes were treated with enhanced chemiluminescence reagents (ECL; Amersham Life Science, Buckinghamshire, UK) and apposed to XAR-5 x-ray film (Eastman Kodak Co., Rochester, NY). Membranes were reprobbed with anti- β -actin antibody as a loading control. In Western experiments to monitor p-CREB, the purity of the nuclear and cytosolic fractions was routinely monitored by reprobing membranes with antihistone H3 antibody (nuclear marker; see Fig. 4C). Whereas histone H3 labeling was strong in the nuclear fractions, it was absent in all cytosolic fractions examined.

To quantitate protein abundance, bands on Western blots were analyzed with a Scan Jet 4-C computing densitometer using NIH IMAGE 1.61 software. Band densities for p-CREB or p-ERK1 and p-ERK2 were corrected for variations in loading and normalized to the corresponding band densities for total CREB or total ERK1 and ERK2, respectively; normalized means were expressed as a percentage of the corresponding value for control (sham operated) animals. Band densities for Bcl-2 were expressed as a percentage of the corresponding value for control animals. Statistical comparisons were assessed by ANOVA, followed by the Newman-Keuls test. Because of the large number of treatment groups, which included two surgical conditions (sham *vs.* ischemia), two hormone treatments (estradiol *vs.* placebo), multiple time points after surgery, and two drugs (PD98059 *vs.* vehicle), it was not always possible to run samples for all conditions on a single gel. Therefore, to enable comparisons from experiment to experiment, band densities for all samples on a given gel were normalized to the band density for a sample from an animal treated with placebo, subjected to sham operation, and killed 1 h after surgery (control). Each gel included at least one sample from such a control animal to enable comparisons of data across different experiments, and a different control animal was prepared for each experiment.

Immunolabeling

Animals were deeply anesthetized with pentobarbital (50 mg/kg, ip) and perfused transcardially with 4% paraformaldehyde in phosphate buffer [0.1 M (pH 7.4)] at 24 and 48 h after ischemia or sham operation ($n = 3$ independent experiments for each time point and treatment group). Brains were removed, postfixed (2 h at 4 C), frozen, and cut into

sections (40 μ m) in the coronal plane of the dorsal hippocampus (3.3–4.0 mm posterior from bregma). Free-floating sections were blocked in 10% normal serum, 5% BSA, and 0.01% saponin in PBS (2 h at room temperature) and processed for immunolabeling with anti-Bcl-2 polyclonal antibody, overnight at 4 C, followed by biotinylated goat antirabbit IgG (1:200; Vector Laboratories, Burlingame, CA). Sections were then incubated with avidin peroxidase complex (ABC kit, 1 h at room temperature; Vector Laboratories), followed by 3–3'-diaminobenzidine (Vector Laboratories). Images were viewed through a Nikon inverted microscope ECLIPSE TE300 and images acquired with a SPOT RT CCD-cooled camera with Diagnostic software version 3.0 (Diagnostic Instruments, Inc., Sterling Heights, MI).

Caspase activity assay

Caspase activity assays were performed on fresh-frozen brain sections using an APO LOGIX™ carboxyfluorescein (FAM) caspase detection kit (Cell Technology, Minneapolis, MN) according to the manufacturer's instructions. FAM-DEVD (Asp-Glu-Val-Asp)-fluoromethyl ketone (FMK) is a carboxy-fluorescein analog of zDEVD (N-benzyloxy-carbonyl-Asp-Glu-Val-Asp)-FMK, a broad-spectrum cysteine protease inhibitor that enters cells and irreversibly binds activated caspases (31–33). FAM-DEVD-FMK exhibits higher affinity for caspase-3 than caspase-8, caspase-7, caspase-10, or caspase-6 (34) and exhibits much lower affinity for the calpains than for caspases; thus, at 5 μ M FAM-DEVD is a relatively selective inhibitor of caspase-3. Moreover, FAM-DEVD-FMK labeling of CA1 neurons correlates well with caspase-3 activation, as assessed by Western blot analysis. In this study we therefore refer to FAM-DEVD-FMK labeling as indicative of caspase-3 activity. In brief, animals were deeply anesthetized with pentobarbital (50 mg/kg, ip) and killed by decapitation at 24 h after ischemia or sham operation (control). Brains were removed, frozen, and cut into sections (18 μ m) in the coronal plane of the dorsal hippocampus. Brain sections (three per animal) were labeled with 5 μ M FAM-DEVD-FMK (1 h, 37 C), washed three times with 1 \times working dilution wash buffer and viewed under a Nikon ECLIPSE TE-300 fluorescent microscope equipped with an image analysis system at an excitation wavelength of 488 nm and emission wavelength of 565 nm. Images were acquired with a SPOT RT CCD-cooled camera with Diagnostic software version 3.0. For quantitation of caspase-3 activity, the fluorescence intensity within the entire hippocampal CA1 cell layer of the images was analyzed using NIH Image 1.61. The mean fluorescence intensity of CA1 in the right and left hemisphere from each of the three sections was averaged to provide a single value for each animal.

Statistical analysis

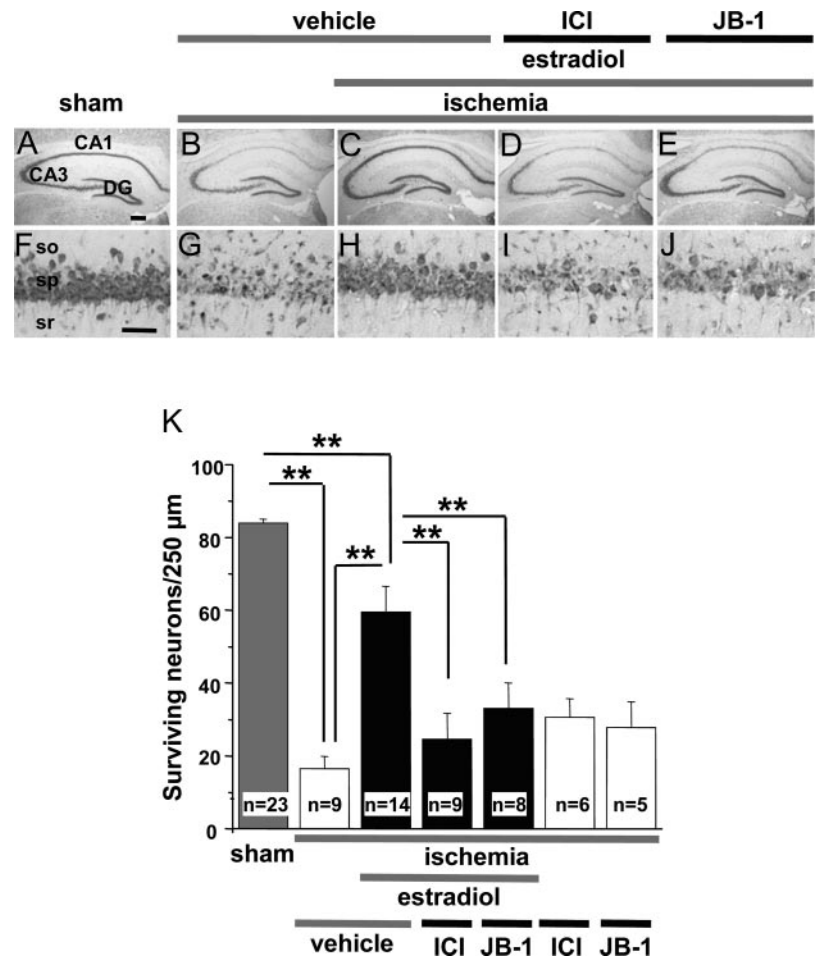
The results were expressed as mean \pm SEM. Data analysis was performed using Pharmacological Calculation System (Springer Verlag, New York, NY; version 4.2). Statistical comparisons were made between groups using a one-way ANOVA followed by Newman-Keuls *post hoc* analysis (neuron counts, immunoblots and caspase-3 activity). T test was used for the serum estradiol data. Differences were considered significant at $P < 0.05$.

Results

Estradiol acts via the classic estrogen receptors and IGF-I receptor to protect CA1 neurons

The intracellular signaling pathways that mediate the neuroprotective actions of estradiol in hippocampal neurons subjected to global ischemia are not well delineated. To address this issue, we treated ovariectomized gerbils with estradiol or placebo for 14 d, subjected animals to sham operation or global ischemia, and administered the broad-spectrum estrogen receptor antagonist ICI 182,780 (100 μ g in 50% DMSO, icv) or vehicle (50% DMSO) into the lateral ventricles at 0 and 12 h after ischemia or sham surgery. Global ischemia induced extensive death of pyramidal cells in the hippocampal CA1 evident at 7 d ($P < 0.01$ *vs.* sham

FIG. 1. The estrogen receptor antagonist ICI 182,780 and IGF-I receptor antagonist JB-1 block estradiol protection. Ovariectomized female gerbils were treated with estradiol or placebo for 14 d and subjected to global ischemia or sham operation. Animals received the estrogen receptor antagonist ICI 182,780 (ICI, 100 μg in 50% DMSO), the IGF-I receptor antagonist JB-1 (10 μg in saline), or vehicle (50% DMSO or saline) icv at 0 and 12 h after ischemia. ICI 182,780 and JB-1 did not affect neuronal survival in ischemic (two rightmost bars in K) or sham animals (supplemental Fig. 1) but greatly reduced estradiol protection, assessed at 7 d after surgery (D, I, E, J, and K). Neither vehicle significantly altered neuronal survival *vs.* no treatment in placebo- or estradiol-treated animals subjected to sham operation. Therefore, vehicle and control data were pooled for purposes of illustration (gray bar labeled sham; see gray bars in supplemental Fig. 1 for different groups). Similarly, neither vehicle significantly altered neuronal survival *vs.* no treatment in placebo-treated animals subjected to global ischemia. Thus, these data were also pooled (white bar labeled ischemia+vehicle; see white bars in supplemental Fig. 1 for different groups). so, Stratum oriens; sp, stratum pyramidale; sr, stratum radiatum. Scale bars, Lower magnification, 400 μm ; higher magnification, 60 μm . **, $P < 0.01$.



animals); the few remaining pyramidal neurons were severely damaged and appeared pyknotic (compare Fig. 1, B and G, with Fig. 1, A, and F; and see Fig. 1K). Qualitative analysis revealed that neuronal death was specific in that little or no cell loss occurred in the nearby CA2 or transition zone or CA3 or dentate gyrus (Fig. 1B). Estradiol reduced the loss of CA1 neurons by approximately 60% ($P < 0.01$ *vs.* ischemia, Fig. 1, C, H, and K). Plasma estradiol levels at the time of death were 15.4 ± 1.4 pg/ml in the placebo group and 136.35 ± 12.9 pg/ml in the estradiol group. The estrogen receptor antagonist ICI 182,780 did not detectably alter the appearance or number of surviving neurons in placebo or estradiol-treated animals subjected to sham surgery (see supplemental Fig. 1, published as supplemental data on The Endocrine Society's Journals Online Web site at <http://endo.endojournals.org>), but ICI 182,780 abrogated the neuroprotective action of estradiol in CA1, assessed at 7 d after ischemia ($P < 0.01$ *vs.* estradiol, Fig. 1, D, I, and K). Moreover, the vehicle for ICI 182,780 had no effect on surviving neurons in either sham-operated or ischemic animals (Fig. 1K and supplemental Fig. 1). These findings indicate that estradiol protection of hippocampal neurons requires activation of the classical estrogen receptors- α and/or - β and is similar to our findings with female rats (30).

To examine a possible role for brain IGF-I receptor activation in estradiol protection of CA1 neurons, we adminis-

tered the highly selective and potent IGF-I receptor antagonist JB-1 (10 μg in saline) or vehicle (saline) icv at 0 and 12 h after reperfusion. JB-1 is a peptide analog of IGF-I corresponding to the recognition motif or D domain within the carboxy terminus of IGF-I that binds the IGF-I receptor. JB-1 effectively blocks IGF-I receptor-mediated autophosphorylation and cellular proliferation (35). JB-1 did not detectably alter the number or appearance of neurons in placebo-treated animals subjected to global ischemia (supplemental Fig. 1) but abolished estradiol protection of CA1 neurons, assessed at 7 d after ischemia ($P < 0.01$ *vs.* estradiol, Fig. 1, E, J, and K). Saline and JB-1 alone had no effect on surviving neurons in either sham or ischemic animals (Fig. 1K; see also Fig. 1 legend and supplemental Fig. 1). These findings indicate that brain IGF-I receptors are critical to estradiol protection of gerbil hippocampal neurons in global ischemia.

MAPK signaling is critical to estradiol protection of CA1 neurons

Because estradiol and IGF-I are known upstream regulators of ERK/MAPK signaling in hippocampal neurons (19, 36), we next examined a possible role for MAPK signaling in estradiol protection. Many of the available antibodies used in the present study recognize rat (but not gerbil) antigens with high affinity and good signal to noise. Thus, these experi-

ments were conducted in rats. Ovariectomized female rats pretreated with estradiol or placebo for 14 d were subjected to global ischemia or sham operation, and the MAPK kinase (MEK) inhibitor PD98059 or vehicle was administered into the lateral ventricle at 0 and 12 h after surgery. Global ischemia induced extensive death of pyramidal cells in the hippocampal CA1 at 7 d after ischemia ($P < 0.01$ vs. sham; compare Fig. 2, D and K, with Fig. 2, A and H; and see Fig. 2O). As observed for gerbils, estradiol did not detectably alter the appearance or number of CA1 neurons in sham-operated rats (Fig. 2, B, I, and O) but greatly reduced the ischemia-induced neuronal loss ($P < 0.01$ vs. ischemia, Fig. 2, F, M, and O). Plasma estradiol levels at the time of death were 20.6 ± 1.3 pg/ml in the placebo group and 58.1 ± 3.8 pg/ml in the estradiol group. The MEK inhibitor PD98059 did not detectably alter the number or appearance of surviving neurons in sham-operated rats (Fig. 2, C, J, and O) but abrogated the neuroprotective action of estradiol in the hippocampal CA1 ($P < 0.01$ vs. estradiol alone, Fig. 2, G, N, and O). In contrast, PD98059 did not affect neuronal survival in placebo-treated animals subjected to sham surgery (Fig. 2, C, J, and O) or ischemia (Fig. 2, E, L, and O). These findings indicate that PD98059 administered icv is neither toxic nor protective in the global ischemia model. Consistent with this, PD98059 does not impair locomotion, and its inhibitory actions on hormone-dependent behaviors are readily reversible (37, 38). Together, these findings indicate that ERK/MAPK signaling

is critical to estradiol protection of hippocampal neurons in a rat model of global ischemia.

Estradiol prevents dephosphorylation and inactivation of ERK1 in postischemic CA1 neurons

Because PD98059 administered at 0 and 12 h after ischemia blocked the protective actions of estradiol, we reasoned that ischemia might inactivate ERK/MAPK in the first few hours after insult. To examine the effects of ischemia and estradiol on MAPK activity, we subjected rats that had been treated with estradiol or placebo for 14 d to global ischemia or sham operation and examined ERK1/2 and p-ERK1/2 abundance in CA1 at 1, 3, and 24 h after reperfusion. Global ischemia significantly reduced phosphorylation of ERK1 in CA1, evident at 1 h after ischemia (reduction to 64% of control; $P < 0.05$; Fig. 3, A and B); at 3 h, p-ERK1 levels were not significantly different from controls (Fig. 3, A and B). Estradiol significantly increased ERK1 and ERK2 phosphorylation in sham-operated animals (ERK1, increase to ~200% of control, $P < 0.01$, Fig. 3, A and B; ERK2, increase to ~142% of control, $P < 0.05$, Fig. 3, A and C) and prevented the early ischemia-induced dephosphorylation of ERK1 (Fig. 3, A and B). Whereas p-ERK1 was reduced at 1 h after ischemia, it was increased significantly in the CA1 at 24 h after ischemia vs. sham-operated animals (Fig. 3D). In contrast, global ischemia did not significantly alter ERK2 phosphorylation at any times examined (Fig. 3, A, C, and E).

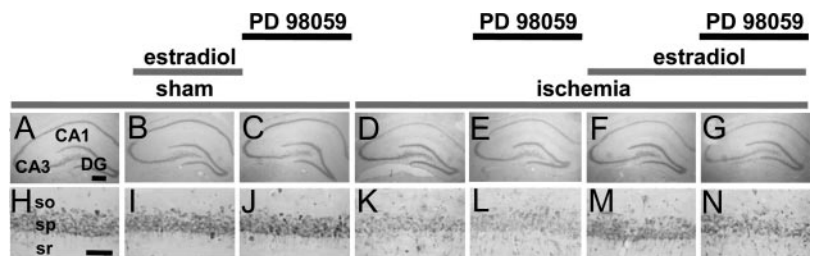
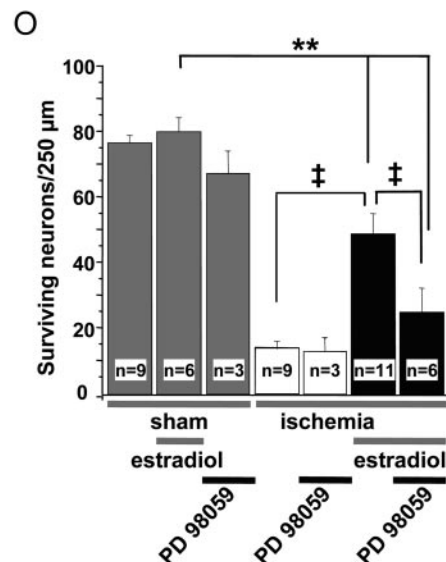


FIG. 2. The ERK/MAPK inhibitor PD98059 attenuates estradiol protection. Ovariectomized female rats were treated with estradiol or placebo for 14 d and subjected to global ischemia (white and black bars) or sham operation (gray bars). Animals received PD98059 (3 μ g) or vehicle (10% DMSO) icv at 0 and 12 h after ischemia. Global ischemia induced extensive death of pyramidal cells in the hippocampal CA1 at 7 d after ischemia ($P < 0.01$ vs. sham). PD98059 greatly reduced estradiol protection, assessed at 7 d after surgery. Abbreviations as in Fig. 1. Scale bars, Lower magnification, 400 μ m; higher magnification, 60 μ m. **, $P < 0.01$ vs. all sham groups; ‡, $P < 0.01$ ischemia+estradiol vs. ischemia+estradiol+PD98059.



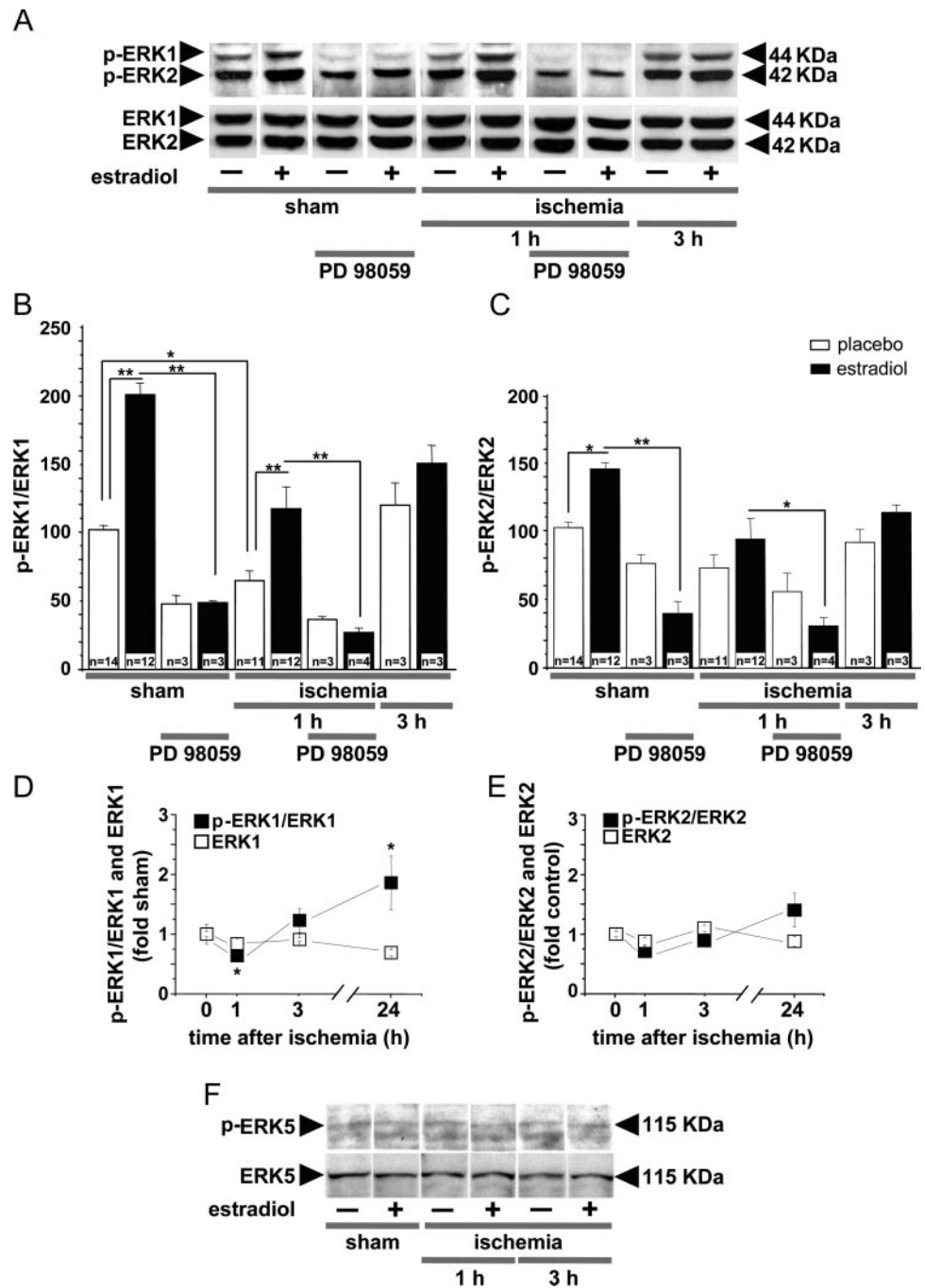


FIG. 3. Estradiol prevents ischemia-induced dephosphorylation of ERK1 in CA1. Representative Western blots (A) and relative abundance of p-ERK1 (B) and p-ERK2 (C) in CA1 whole-cell lysates from placebo- and estradiol-treated rats subjected to sham surgery or ischemia at 1, 3, or 24 h after surgery. Subsets of animals were injected with PD98059 at 0 h and killed at 1 h after surgery. Westerns were probed with antibodies to p-ERK1/2 and ERK1/2. Ischemia-induced dephosphorylation of ERK1 (A and B) but not ERK2 (A and C). Estradiol significantly enhanced ERK1 (B) and ERK2 (C) phosphorylation in shams and maintained levels of p-ERK1 (B) in ischemic rats. PD98059 blocked the estradiol-induced increase in phosphorylation of ERK1 (A and B) in control (sham) CA1. PD98059 also reversed the ability of estradiol to maintain ERK1 phosphorylation in post-ischemic CA1 (A and B). ERK1 dephosphorylation was maximal at 1 h after ischemia; by 24 h, ERK1 phosphorylation was increased relative to that of sham animals (D). In the absence of estradiol treatment, ERK2 and p-ERK2 levels were unchanged at all times examined after ischemia (E). Neither ischemia nor estradiol treatment altered levels of ERK5 or p-ERK5 in CA1 at 1 or 3 h after ischemia (F); ERK5 data are representative of four to five animals per group. For B–E, values for ischemic rats were normalized to the corresponding sham value. *, $P < 0.05$; **, $P < 0.01$.

Because PD98059 at high concentrations can also inhibit MEK5 and activation of ERK5 (39), we assessed whether ERK5 phosphorylation was modified by ischemia or estradiol. Neither global ischemia nor estradiol significantly altered abundance or phosphorylation of ERK5 at 1 or 3 h after insult (Fig. 3F). These findings indicate that global ischemia induces dephosphorylation of ERK1 (but not ERK2 or 5) in the early postischemic period and that estradiol maintains p-ERK1 at higher levels than would otherwise be observed in the postischemic period. PD98059 administered immediately after reperfusion blocked the estradiol-induced increase in phosphorylation of ERK1 (Fig. 3, A and B) and ERK2

(Fig. 3, A and C) in sham-operated animals and reversed the ability of estradiol to maintain ERK1 phosphorylation in postischemic CA1, assessed at 1 h after surgery (Fig. 3, A and B). These findings demonstrate that PD98059 infused into the ventricles reaches the hippocampal CA1 and effectively inhibits its target MEK1/2.

Estradiol prevents ischemia-induced dephosphorylation and inactivation of CREB in CA1

A well-characterized downstream target of ERK/MAPK signaling is the transcription factor CREB, which promotes

transcription of a number of prosurvival genes. ERK/MAPK rapidly induces sustained phosphorylation of CREB at serine 133 (40). To examine whether estradiol regulates phosphorylation and activation of CREB, we subjected estradiol- and placebo-treated rats to global ischemia or sham operation and examined CREB and p-CREB abundance in CA1 at 1 and 3 h after reperfusion. Global ischemia induced a significant decrease in p-CREB, with no significant change in total CREB abundance in the nuclear fraction of CA1 (decrease by ~25% at 1 h, $P < 0.05$ vs. sham-operated animals; decrease by ~40% at 3 h; $P < 0.01$ vs. sham-operated animals, Fig. 4, A and B). Estradiol prevented the ischemia-induced dephosphorylation and inactivation of CREB, evident at both 1 and 3 h after ischemia ($P < 0.05$ vs. placebo-treated, ischemic animals at 1 h, $P < 0.01$ vs. placebo-treated ischemic animals at 3 h; Fig. 4, A and B). Thus, estradiol blocks insult-induced dephosphorylation and inactivation of CREB in the vulnerable CA1. PD98059 administered immediately after ischemia blocked the ability of estradiol to maintain p-CREB levels in post-ischemic CA1, assessed at 3 h after surgery. These findings suggest a causal role for ERK/MAPK signaling in the ability of estradiol to maintain CREB phosphorylation and activation in postischemic CA1.

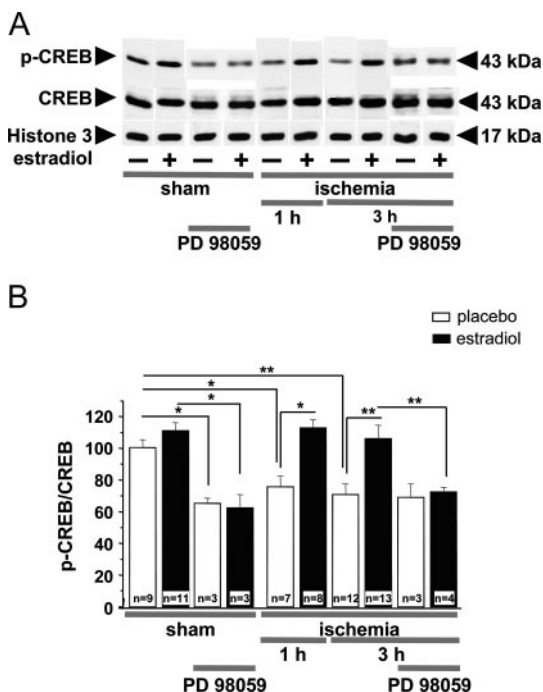


FIG. 4. Estradiol acts via MAPK signaling to prevent ischemia-induced CREB dephosphorylation. Representative Western blots (A) and relative p-CREB abundance (B) in the nuclear fraction of CA1 of placebo- and estradiol-treated rats subjected to sham operation or ischemia at 1 and 3 h after surgery. Subsets of animals were injected with PD98059 at 0 h and killed at 3 h after surgery. Westerns were probed with antibodies to p-CREB, CREB, and histone 3 (nuclear marker). Ischemia promoted dephosphorylation of CREB in the nuclei of CA1 neurons. Estradiol maintained the phosphorylated, activated state of CREB in the CA1 of ischemic rats. PD98059 blocked the ability of estradiol to maintain p-CREB levels in postischemic CA1. Values for experimental animals were normalized to the corresponding sham value. **, $P < 0.01$; *, $P < 0.05$.

Estradiol attenuates ischemia-induced decrease of Bcl-2 protein expression in CA1 neurons

Bcl-2 is an antiapoptotic factor implicated in preservation of the integrity of the mitochondrial outer membrane, and the *bcl-2* gene is a downstream target of estradiol (41, 42) and CREB (for review, see Refs. 43 and 44). Estradiol can also act indirectly to regulate Bcl-2 gene expression by promoting CREB phosphorylation (42, 45). To examine the effects of estradiol pretreatment and ischemia on Bcl-2 protein expression, we subjected estradiol- and placebo-treated animals to global ischemia or sham operation and examined Bcl-2 abundance in CA1 at later times (12, 24, or 48 h) after reperfusion. Global ischemia induced a marked decrease in Bcl-2 expression in pyramidal neurons of the hippocampal CA1, evident at 24 and 48 h, times before onset of neuronal death, as revealed by immunolabeling (Fig. 5, E–H). Estradiol enhanced Bcl-2 immunolabeling in CA1 neurons of sham-operated animals (Fig. 5, A–D) and blocked the ischemia-induced down-regulation of Bcl-2 in CA1 neurons of ischemic animals (Fig. 5, I–L). Because the loss of CA1 pyramidal neurons in rats subjected to 10 min of global ischemia is not detectable until 2–3 d after ischemia (26), it is unlikely that the reduced Bcl-2 immunolabeling at 12 and 24 h simply reflects pyramidal cell death. To assess the effects of ischemia and estradiol on Bcl-2 protein abundance, protein samples from the CA1 were subjected to Western blot analysis (Fig. 5M). Global ischemia induced a marked (~50%) decrease in Bcl-2 protein abundance in CA1, evident at 12 h ($P < 0.05$ vs. placebo-treated, sham animals). Bcl-2 was modestly, but not significantly, decreased as late as 24 and 48 h after ischemia (Fig. 5, M and N). Estradiol blocked the ischemia-induced down-regulation of Bcl-2 in CA1 at 12 h ($P < 0.05$ vs. ischemia at 12 h), with no detectable decline in Bcl-2 as late as 24 or 48 h after reperfusion (Fig. 5, M and N).

Because the MAPK inhibitor PD98059 blocks the neuroprotective actions of estradiol, we next determined whether ERK/MAPK signaling is causally related to the effects of estradiol on Bcl-2 protein abundance. PD98059 administered immediately after ischemia did not prevent estradiol maintenance of Bcl-2 abundance in CA1 (Fig. 5, M and N), even though this drug blocked the ability of estradiol to promote CA1 pyramidal cell survival (Fig. 2). Interestingly, PD98059 alone significantly increased Bcl-2 in the CA1 of estradiol- (but not placebo) treated sham and ischemic animals (Fig. 5, M and N). Therefore, whereas estradiol appears to maintain CREB signaling in the postischemic CA1 via the ERK/MAPK pathway (Fig. 4), the positive effects of estradiol on Bcl-2 expression are evident, even when this pathway is blocked, and the effects of estradiol and PD98059 on Bcl-2 may be additive. Hence, our findings are consistent with a model whereby estradiol acts on the Bcl-2 gene in a MAPK-independent manner to sustain Bcl-2 gene expression at normal levels in postischemic CA1 neurons.

Estradiol blocks ischemia-induced activation of caspase-3 activity in CA1 neurons

Injurious stimuli such as global ischemia disrupt the integrity of the mitochondrial membrane, leading to the release of cytochrome *c* and activation of caspase-3, a terminator

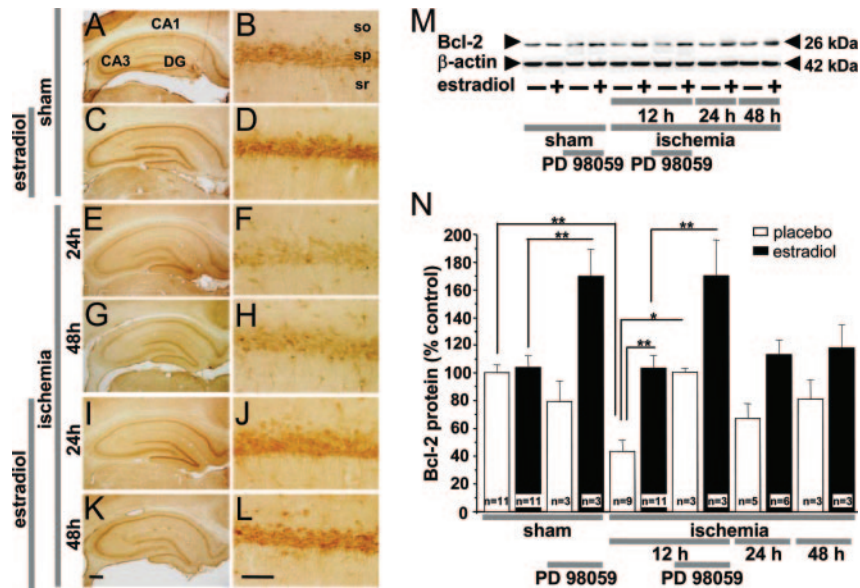


FIG. 5. Estradiol prevents ischemia-induced down-regulation of Bcl-2 in CA1. Bcl-2 immunolabeling in the CA1 pyramidal cell layer in brain sections at the level of the dorsal hippocampus from ovariectomized rats subjected to sham operation (A–D) or global ischemia at 24 h (E and F and I and J) and 48 h (G and H and K and L) after reperfusion. Data are typical of a minimum of three animals per time point and treatment group. Estradiol attenuated the ischemia-induced decrease in Bcl-2 in CA1 neurons. Abbreviations as in Fig. 1. Scale bars, Lower magnification, 400 μ m; higher magnification, 60 μ m. Representative Western blots (M) and relative abundance of Bcl-2 (N) in the cytosolic fraction of CA1 from placebo- and estradiol-treated rats subjected to sham operation or global ischemia at 12, 24, and 48 h after surgery. Subsets of animals were injected with PD98059 at 0 h and killed at 12 h after surgery. Westerns were probed with antibodies to Bcl-2 and β -actin (loading control). Global ischemia caused down-regulation of Bcl-2 at 12 h. Estradiol prevented the ischemia-induced down-regulation of Bcl-2. PD98059 did not block the ability of estradiol to maintain Bcl-2 levels, and increased Bcl-2 abundance in the CA1 of estradiol-treated sham and ischemic animals (M and N). Band densities for experimental animals were normalized to the corresponding values for control animals. *, $P < 0.05$; **, $P < 0.01$.

caspace implicated in the execution step of apoptosis (for review, see Ref. 1). Global ischemia promotes cleavage of the biologically inactive precursor procaspase-3 to generate activated caspase-3 (46); ischemia-induced caspase-3 activity is maximal at 24 h after insult (47). To directly measure caspase-3 functional activity after ischemia, we labeled brain sections with FAM-DEVD-FMK, a fluorescein-tagged analog of the caspase inhibitor zDEVD-FMK, at 24 h. FAM-DEVD-FMK enters cells and binds irreversibly to catalytically active caspase-3 and thus provides a fluorescent indicator of the abundance of active caspase-3. In brain sections from control animals, caspase activity was low (Fig. 6, A and B). Global ischemia induced a dramatic, 6-fold increase in caspase activity in the hippocampal CA1, evident at 24 h ($P < 0.01$ vs. placebo-treated, sham animals; Fig. 6, C, D, and G). The increase in caspase activity was specific in that it was not observed in the resistant CA3 or dentate gyrus. Estradiol pretreatment (14 d) completely blocked the ischemia-induced elevation of caspase-3 activity in CA1 (Fig. 6, E–G).

Discussion

Estradiol at levels used for hormone therapy in women ameliorates hippocampal injury in animals subjected to global ischemia (46, 48, 49). Here we report the novel observations that estradiol acts *in vivo* via classical estrogen receptors, IGF-I receptors, and ERK/MAPK signaling to promote survival of postischemic hippocampal CA1 neurons in ovariectomized rodents. Ischemia promotes dephosphorylation and inactivation of ERK1 and the transcription factor CREB (a known ERK target) in the first few hours after

ischemia, subsequent down-regulation of the antiapoptotic protein Bcl-2 and activation of caspase-3. Estradiol prevents ischemia-induced dephosphorylation and inactivation of ERK1 and CREB, down-regulation of Bcl-2, and caspase activation. ERK/MAPK signaling is critical to CREB activation and neuronal survival in that administration of the MEK inhibitor PD98059 blocks the ability of estradiol to maintain ERK1 and CREB phosphorylation and to promote survival of CA1 pyramidal neurons in the postischemic hippocampus. In contrast, the impact of estradiol on Bcl-2 abundance is ERK independent, consistent with the concept that the activated estrogen receptor interacts with enhancers such as cAMP response elements or specificity protein-1 sites on the *bcl-2* gene to regulate *bcl-2* gene expression (see Ref. 42). These findings support a model whereby estradiol acts via classical estrogen receptors and IGF-I receptors (Fig. 7), which converge on ERK/MAPK signaling and CREB to promote neuronal survival in the face of ischemic insults. Although estradiol neuroprotection has previously been shown to require concurrent IGF-I signaling in other animal models (*e.g.* see Refs. 19 and 20), a recent clinical study demonstrating a positive correlation between the level of circulating IGF-I and better functional outcomes in ischemic stroke suggests that adequate levels of IGF-I receptor activity may be relevant to the human situation (see Ref. 50).

Whereas estradiol neuroprotection in animal models of focal and global ischemia is well established (12–15 but also see Ref. 51), the therapeutic potential of estrogens in ameliorating the sequelae of cardiac arrest (global ischemia) or stroke (focal ischemia) in humans remains controversial. The

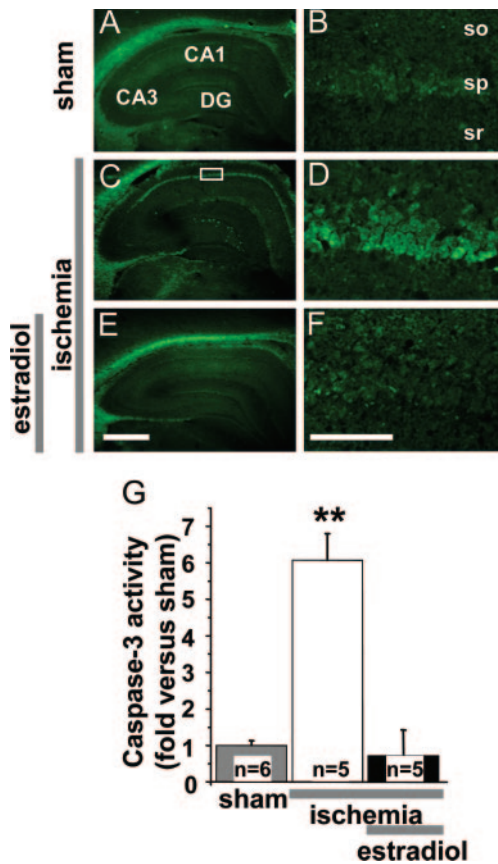


FIG. 6. Estradiol blocks ischemia-induced caspase-3 activity in CA1. Representative brain sections at the level of the dorsal hippocampus from control (A and B) and experimental animals subjected to global ischemia (C–F) labeled with FAM-DEVD-FMK, a cell-permeant, irreversible inhibitor of caspase-3. In control brain caspase-3 activity was undetectable (A, B, and G). Global ischemia activated caspase-3 in CA1 neurons, evident at 24 h (C, D, and G). Estradiol prevented ischemia-induced caspase-3 activity (E–G). Abbreviations as in Fig. 1. Scale bars, Lower magnification, 400 μ m; higher magnification, 50 μ m. **, $P < 0.01$ vs. sham and ischemia+estradiol.

Women’s Health Initiative Memory Study indicates that hormone treatment initiated many years after menopause does not protect against dementia or cognitive decline in elderly women (52, 53). Moreover, postmenopausal treatment with estrogen plus progestin impairs global cognitive performance in some subjects (53). One explanation for these unanticipated outcomes might be the long (years) duration of the interval between cessation of ovarian function and initiation of hormone therapy (see Ref. 54); therefore, the significance of carefully controlled estradiol studies in animals has increased (55).

Estrogen receptor/IGF-I receptor interactions in estradiol neuroprotection

Our study provides new insight into intracellular signaling pathways that mediate estradiol neuroprotection in an *in vivo* model of global ischemia. The estrogen receptor antagonist ICI 182,780 abrogated estradiol protection in gerbils (present study) and rats (30), documenting a critical role for classical estrogen receptors in protection of hippocampal

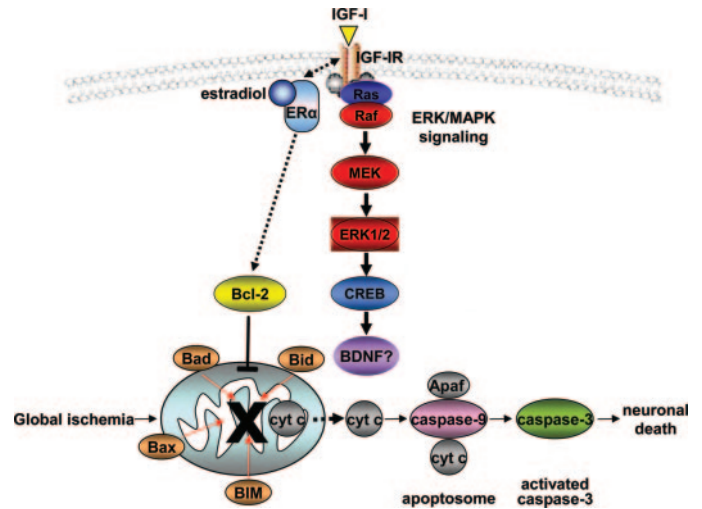


FIG. 7. Hypothesized scheme of estradiol protection against global ischemia-induced cell death. Upon stimulation with estradiol, the estrogen receptor (ER)- α [not ER β (23)] forms a macromolecular signaling complex with the IGF-I receptor (IGF-IR) in the plasma membrane; the complex recruits and activates downstream kinases including ERK/MAPK (24, 25). ERK/MAPK signaling culminates in phosphorylation and activation of nuclear transcription factors such as CREB, which regulate target genes important to neuronal survival. CREB targets implicated in neuronal survival include BDNF and the antiapoptotic protein Bcl-2 (43). Bcl-2 promotes the integrity of the mitochondrial outer membrane and halts the caspase death cascade, enabling neurons to survive. Estradiol maintains ERK and CREB activation and opposes ischemia-induced down-regulation of Bcl-2. Data in the present study are consistent with a direct action of estradiol on Bcl-2 gene expression rather than maintenance of Bcl-2 levels by ERK/MAPK and CREB signaling. Apaf, Apoptotic protease activating factors; Bax, Bcl-2 associated X protein; BIM, Bcl-2 interacting mediator of cell death; Bad, Bcl-2 associated death protein; Bid, BH3 interacting domain death agonist; cyt c, cytochrome c.

neurons from global ischemia. The fact that both estrogen receptor- α and - β -selective agonists produce neuroprotection (30, 49) suggests that either receptor can mediate estradiol protection in global ischemia. In contrast, studies involving mice with targeted gene deletions indicate that estrogen receptor- α , but not - β , mediates estradiol protection in focal ischemia (56). Moreover, we cannot rule out a role for G protein-coupled receptor (GPR)-30, a G protein-coupled receptor implicated in estradiol regulation of epidermal growth factor receptor signaling. GPR30 binds estradiol with nanomolar affinity, and ICI 182,780 inhibits estradiol signaling mediated by GPR30 (57, 58). Thus, estradiol might promote neuronal survival by different mechanisms in the global and focal ischemia paradigms.

The finding that the competitive IGF-I receptor antagonist JB-1 also blocks estradiol protection documents a role for IGF-I receptor signaling in preservation of hippocampal neurons after global ischemia. This finding is consistent with evidence that estrogen receptor- α and IGF-I receptors are functionally linked (24, 25, 59). Activation of estrogen/IGF-I signaling is implicated in protection of hilar neurons from seizure-induced injury (60), protection of retinal neurons against light-induced degeneration (61), synaptic remodeling of hypothalamic neurons *in vivo* (62), and estradiol-

dependent reproductive behaviors and neuroendocrine responses (63). Recent studies have begun to reveal the molecular underpinnings of this interdependence. Estrogen receptor- α and IGF-I receptors form a macromolecular signaling complex that recruits downstream signaling molecules such as Shc, phosphatidylinositol 3-kinase and insulin receptor substrate-1 (23). Although estrogen receptors also interact with epidermal growth factor receptors and HER-2/neu in peripheral reproductive tissues and tumor cells (57, 64), the relevance to neurons is unclear.

The present experiments were performed in rats, with the exception of ICI and JB-1 experiments, which were in gerbils. Although not confirmed in rats, a role for IGF-I receptors in estradiol protection against ischemic damage is likely to apply across species. First, experiments using JB-1 show that IGF-I receptors are critical to estradiol protection against seizure-induced damage of hippocampal neurons (20) and light-induced damage of retinal neurons in rats (61). Second, there are strong commonalities between the gerbil and rat models of global ischemia (1). In both species, cell death occurs primarily in the hippocampal CA1, in which pyramidal neurons (but not interneurons or astrocytes) die. In both species, histological evidence of cell death is not detectable until at least 48 h. Moreover, ischemia promotes activation of the same death cascades, translocation of the same proteins, and alterations in expression of the same genes, albeit with a somewhat slower time course in gerbils (26, 27, 47). Finally, it is unlikely that there are species differences in the actions of the IGF-I antagonist JB-1, which was developed to target human IGF-I receptors (35).

ERK/MAPK signaling is critical to neuronal survival

ERK/MAPK signaling is a well-known downstream target of estradiol and IGF-I in neurons (65). We show that estradiol pretreatment promotes phosphorylation and activation of ERK1/2 in CA1 of sham-operated animals and blocks ischemia-induced dephosphorylation of ERK1. Furthermore, administration of the ERK/MAPK antagonist PD98059 after ischemia reverses estradiol-dependent ERK1/2 phosphorylation and neuroprotection. These findings implicate ERK/MAPK signaling in protection of hippocampal neurons and are consistent with observations that ERK/MAPK mediates estradiol neuroprotection in models of quinolinic acid (66) and glutamate excitotoxicity (67), β -amyloid toxicity (68), and oxidative stress (69). In contrast, PD98059 protects neurons in focal ischemia (70–72). Thus, ERK/MAPK signaling can promote neuronal apoptosis in some cases (73).

Although the PD98059 experiments suggest a role for ERK1 signaling in estradiol protection, we cannot rule out a role for other targets of PD98059 such as MEK5 and/or cyclooxygenase (39). However, we believe that these molecules are unlikely to mediate estradiol neuroprotection. First, estradiol clearly promotes phosphorylation and activation of ERK1/2, but not ERK5. Second, PD98059 reverses the effects of estradiol on ERK1/2 phosphorylation in both sham-operated and ischemic females. Third, PD98059 exhibits a higher affinity for MEK1/2 than for MEK5; thus, at the concentration used in the present study PD98059 may not inhibit ERK5. Moreover, ischemia does not alter ERK5 abundance or

activity in CA1 (Ref. 74, present study), providing strong evidence for target specificity. Fourth, although estradiol regulates cyclooxygenase-2 expression, it does not regulate enzyme activity (75). Finally, PD98059 is a standard pharmacological agent for inhibiting ERK1/2 signaling in neurons *in vivo* and *in vitro* (76–80). Behavioral studies involving PD98059 and another ERK inhibitor (U0126) report essentially identical results (37, 76). Whereas our findings implicate ERK/MAPK signaling as a necessary cellular mediator of estradiol protection, this pathway is not necessarily sufficient for neuronal survival. In particular, our results do not rule out an important role for phosphatidylinositol 3-kinase, another downstream mediator of estradiol/IGF-I protection (59, 65, 81).

ERK/MAPK signaling is critical to estradiol-dependent CREB activation

The ERK/MAPK cascade can promote neuronal survival by a number of mechanisms including phosphorylation and activation of CREB, which drives transcription of pro-survival genes such as BDNF; phosphorylation and inactivation of the proapoptotic protein Bcl-2-associated death protein (40); and activation of glycogen synthase kinase-3 β , which inhibits Wnt signaling (82). Present findings that estradiol maintains CREB phosphorylation at serine 133 in the face of ischemia and that PD98059 reverses estradiol-dependent CREB phosphorylation indicate a causal relation between ERK/MAPK signaling and CREB activation. Estradiol binding to estrogen receptor- α can also activate CREB by promoting metabotropic glutamate receptor-mediated stimulation of MAPK (83) or inducing cAMP synthesis (42, 45).

ERK/MAPK signaling is not critical to estradiol-dependent regulation of Bcl-2 expression

Our finding that estradiol maintains levels of the antiapoptotic protein Bcl-2 after global ischemia is consistent with studies implicating Bcl-2 in estradiol protection in focal ischemia (84, 85) and *in vitro* models of cell death (67, 69). Although these paradigms target different neurons and induce death via different mechanisms, regulation of Bcl-2 may be a common mechanism of estradiol neuroprotection. However, it is clear that maintaining Bcl-2 is not sufficient to rescue CA1 neurons from ischemia-induced cell death. We found that the MEK inhibitor PD98059 potentiated the effects of estradiol on Bcl-2 expression but completely blocked the effects of estradiol on cell survival and p-ERK1. Estradiol can increase *bcl-2* gene expression by CREB activation in other systems (42, 45), but our finding that PD98059 does not reverse (and may even potentiate) regulation of Bcl-2 levels by estradiol is consistent with a MAPK-independent action of estradiol on *bcl-2* gene expression. The loss of CA1 neurons in rats subjected to 10 min of global ischemia is not detectable until 2–3 d after ischemia (26); therefore, it is unlikely that the reduced Bcl-2 immunoreactivity detected at 12 and 24 h after ischemia simply reflects neuronal cell death. However, it is possible that cell death could contribute to the reduction of Bcl-2 observed at 48 h.

Injurious stimuli such as global ischemia disrupt the functional integrity of the outer mitochondrial membrane, re-

leasing cytochrome *c* and Smac/DIABLO (second mitochondria-derived activator of caspase/direct inhibitor of apoptosis-binding protein with low pI), which activate the caspase death cascade (47, 86–88). Proapoptotic Bcl-2 family members are thought to participate in formation and activation of large, multiconductance channels that mediate cytochrome *c* release (1). Antiapoptotic family members such as Bcl-2 preserve the mitochondrial integrity and block caspase activation in the face of injury. Present findings suggest that estradiol promotes neuronal survival, at least in part, by sustaining levels of Bcl-2. Accordingly, acute estradiol prevents ischemia-induced translocation of cytochrome *c* from mitochondria to the cytosol (89). However, the current findings also suggest that maintenance of Bcl-2 expression is not sufficient to mediate estradiol protection of CA1 neurons because Bcl-2 protein abundance was sustained in estradiol-treated, ischemic females injected with the MEK inhibitor PD98059. Therefore, it is likely that estradiol protects neurons from apoptotic death by regulating the activity of multiple cell survival pathways and that interference with any one of these signaling pathways may lead to delayed cell death.

Summary

The present study shows that estradiol acts via estrogen receptors, IGF-I receptors, and ERK/MAPK signaling to promote CA1 neuronal survival after global ischemia. Ischemia promotes early dephosphorylation and inactivation of ERK1 and its target CREB, followed by down-regulation of Bcl-2 and caspase activation. Estradiol attenuates dephosphorylation of ERK1 and CREB, down-regulation of Bcl-2, and activation of caspase-3. These findings identify intracellular signaling pathways that mediate estradiol protection and reveal mechanisms by which estradiol protects hippocampal neurons from global ischemia-induced degeneration. These findings suggest that estradiol and IGF-I act in a coordinate manner to promote activation of ERK/MAPK signaling and its target CREB and thereby promote neuronal survival in the face of ischemic insults.

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References

- Zukin RS, Jover T, Yokota H, Calderone A, Simionescu M, Lau CG 2004 Molecular and cellular mechanisms of ischemia-induced neuronal death. In: Mohr JP, Choi D, Grotta JC, Weir B, Wolf PA, eds. *Stroke: pathophysiology, diagnosis, and management*. Orlando, FL: Harcourt; 829–854
- Chen J, Zhu RL, Nakayama M, Kawaguchi K, Jin K, Stetler RA, Simon RP, Graham SH 1996 Expression of the apoptosis-effector gene, *Bax*, is up-regulated in vulnerable hippocampal CA1 neurons following global ischemia. *J Neurochem* 67:64–71
- Pulsinelli WA, Brierley JB, Plum F 1982 Temporal profile of neuronal damage in a model of transient forebrain ischemia. *Ann Neurol* 11:491–498
- Rosenbaum DM, D'Amore J, Llena J, Rybak S, Balkany A, Kessler JA 1998 Pretreatment with intraventricular aurointricarboxylic acid decreases infarct size by inhibiting apoptosis following transient global ischemia in gerbils. *Ann Neurol* 43:654–660
- Kirino T 1982 Delayed neuronal death in the gerbil hippocampus following ischemia. *Brain Res* 239:57–69
- Murphy DD, Segal M 1996 Regulation of dendritic spine density in cultured rat hippocampal neurons by steroid hormones. *J Neurosci* 16:4059–4068
- Pozzo-Miller LD, Inoue T, Murphy DD 1999 Estradiol increases spine density and NMDA-dependent Ca²⁺ transients in spines of CA1 pyramidal neurons from hippocampal slices. *J Neurophysiol* 81:1404–1411
- Woolley CS, McEwen BS 1994 Estradiol regulates hippocampal dendritic spine density via an *N*-methyl-D-aspartate receptor-dependent mechanism. *J Neurosci* 14:7680–7687
- Gazzaley AH, Weiland NG, McEwen BS, Morrison JH 1996 Differential regulation of NMDAR1 mRNA and protein by estradiol in the rat hippocampus. *J Neurosci* 16:6830–6838
- Moss RL, Gu Q 1999 Estrogen: mechanisms for a rapid action in CA1 hippocampal neurons. *Steroids* 64:14–21
- Gu Q, Moss RL 1998 Novel mechanism for non-genomic action of 17 β -oestradiol on kainate-induced currents in isolated rat CA1 hippocampal neurones. *J Physiol* 506(Pt 3):745–754
- Amantea D, Russo R, Bagetta G, Corasaniti MT 2005 From clinical evidence to molecular mechanisms underlying neuroprotection afforded by estrogens. *Pharmacol Res* 52:119–132
- Behl C 2002 Oestrogen as a neuroprotective hormone. *Nat Rev Neurosci* 3:433–442
- Garcia-Segura LM, Azcoitia I, DonCarlos LL 2001 Neuroprotection by estradiol. *Prog Neurobiol* 63:29–60
- Wise P 2003 Estradiol exerts neuroprotective actions against ischemic brain injury: insights derived from animal models. *Endocrine* 21:11–15
- Gulinello M, Lebesgue D, Jover-Mengual T, Zukin RS, Etgen AM 2006 Acute and chronic estradiol treatments reduce memory deficits induced by transient global ischemia in female rats. *Horm Behav* 49:246–260
- Plamondon H, Morin A, Charron C 2006 Chronic 17 β -estradiol pretreatment and ischemia-induced hippocampal degeneration and memory impairments: a 6-month survival study. *Horm Behav* 50:361–369
- McEwen B 2002 Estrogen actions throughout the brain. *Recent Prog Horm Res* 57:357–384
- Cardona-Gomez GP, Mendez P, DonCarlos LL, Azcoitia I, Garcia-Segura LM 2002 Interactions of estrogen and insulin-like growth factor-I in the brain: molecular mechanisms and functional implications. *J Steroid Biochem Mol Biol* 83:211–217
- Azcoitia I, Sierra A, Garcia-Segura LM 1999 Neuroprotective effects of estradiol in the adult rat hippocampus: interaction with insulin-like growth factor-I signalling. *J Neurosci Res* 58:815–822
- Sweatt JD 2004 Mitogen-activated protein kinases in synaptic plasticity and memory. *Curr Opin Neurobiol* 14:311–317
- Thomas GM, Haganir RL 2004 MAPK cascade signalling and synaptic plasticity. *Nat Rev Neurosci* 5:173–183
- Kahlert S, Nuedling S, van Eickels M, Vetter H, Meyer R, Grohe C 2000 Estrogen receptor alpha rapidly activates the IGF-1 receptor pathway. *J Biol Chem* 275:18447–18453
- Mendez P, Azcoitia I, Garcia-Segura LM 2003 Estrogen receptor α forms estrogen-dependent multimolecular complexes with insulin-like growth factor receptor and phosphatidylinositol 3-kinase in the adult rat brain. *Brain Res Mol Brain Res* 112:170–176
- Song RX, Barnes CJ, Zhang Z, Bao Y, Kumar R, Santen RJ 2004 The role of Shc and insulin-like growth factor 1 receptor in mediating the translocation of estrogen receptor alpha to the plasma membrane. *Proc Natl Acad Sci USA* 101:2076–2081
- Calderone A, Jover T, Mashiko T, Noh KM, Tanaka H, Bennett MV, Zukin RS 2004 Late calcium EDTA rescues hippocampal CA1 neurons from global ischemia-induced death. *J Neurosci* 24:9903–9913
- Calderone A, Jover T, Noh K-M, Tanaka H, Yokota H, Lin Y, Grooms S, Regis R, Bennett MV, Zukin RS 2003 Ischemic insults de-repress the gene silencer REST in neurons destined to die. *J Neurosci* 23:2112–2121
- Blevins JE, Stanley BG, Reidelberger RD 2002 DMSO as a vehicle for central injections: tests with feeding elicited by norepinephrine injected into the paraventricular nucleus. *Pharmacol Biochem Behav* 71:277–282
- Paxinos GA, Watson C 1998 *The rat brain in stereotaxic coordinates*. San Diego: Academic Press
- Miller NR, Jover T, Cohen HW, Zukin RS, Etgen AM 2005 Estrogen can act via estrogen receptor α and β to protect hippocampal neurons against global ischemia-induced cell death. *Endocrinology* 146:3070–3079
- Amstad PA, Yu G, Johnson GL, Lee BW, Dhawan S, Phelps DJ 2001 Detection

- of caspase activation *in situ* by fluorochrome-labeled caspase inhibitors. *Bio-techniques* 31:608–610, 612, 614, *passim*
32. **Bedner E, Smolewski P, Amstad P, Darzynkiewicz Z** 2000 Activation of caspases measured *in situ* by binding of fluorochrome-labeled inhibitors of caspases (FLICA): correlation with DNA fragmentation. *Exp Cell Res* 259: 308–313
 33. **Smolewski P, Bedner E, Du L, Hsieh TC, Wu JM, Phelps DJ, Darzynkiewicz Z** 2001 Detection of caspases activation by fluorochrome-labeled inhibitors: multiparameter analysis by laser scanning cytometry. *Cytometry* 44:73–82
 34. **Garcia-Calvo M, Peterson EP, Leiting B, Ruel R, Nicholson DW, Thornberry NA** 1998 Inhibition of human caspases by peptide-based and macromolecular inhibitors. *J Biol Chem* 273:32608–32613
 35. **Pietrzkowski Z, Wernicke D, Porcu P, Jameson BA, Baserga R** 1992 Inhibition of cellular proliferation by peptide analogues of insulin-like growth factor 1. *Cancer Res* 52:6447–6451
 36. **Bi R, Broutman G, Foy MR, Thompson RF, Baudry M** 2000 The tyrosine kinase and mitogen-activated protein kinase pathways mediate multiple effects of estrogen in hippocampus. *Proc Natl Acad Sci USA* 97:3602–3607
 37. **Etgen AM, Acosta-Martinez M** 2003 Participation of growth factor signal transduction pathways in estradiol facilitation of female reproductive behavior. *Endocrinology* 144:3828–3835
 38. **Gonzalez-Flores O, Shu J, Camacho-Arroyo I, Etgen AM** 2004 Regulation of lordosis by cyclic 3',5'-guanosine monophosphate, progesterone, and its 5 α -reduced metabolites involves mitogen-activated protein kinase. *Endocrinology* 145:5560–5567
 39. **English JM, Cobb MH** 2002 Pharmacological inhibitors of MAPK pathways. *Trends Pharmacol Sci* 23:40–45
 40. **Bonni A, Brunet A, West AE, Datta SR, Takasu MA, Greenberg ME** 1999 Cell survival promoted by the Ras-MAPK signaling pathway by transcription-dependent and -independent mechanisms. *Science* 286:1358–1362
 41. **Garcia-Segura LM, Cardona-Gomez P, Naftolin F, Chowen JA** 1998 Estradiol upregulates Bcl-2 expression in adult brain neurons. *Neuroreport* 9:593–597
 42. **Dong L, Wang W, Wang F, Stoner M, Reed JC, Harigai M, Samudio I, Kladde MP, Vyhldal C, Safe S** 1999 Mechanisms of transcriptional activation of bcl-2 gene expression by 17 β -estradiol in breast cancer cells. *J Biol Chem* 274:32099–32107
 43. **Polster BM, Fiskum G** 2004 Mitochondrial mechanisms of neural cell apoptosis. *J Neurochem* 90:1281–1289
 44. **Yuan J, Lipinski M, Degtrev A** 2003 Diversity in the mechanisms of neuronal cell death. *Neuron* 40:401–413
 45. **Kanda N, Watanabe S** 2003 17 β -Estradiol inhibits oxidative stress-induced apoptosis in keratinocytes by promoting Bcl-2 expression. *J Invest Dermatol* 121:1500–1509
 46. **Jover T, Tanaka H, Calderone A, Oguro K, Bennett MV, Etgen AM, Zukin RS** 2002 Estrogen protects against global ischemia-induced neuronal death and prevents activation of apoptotic signaling cascades in the hippocampal CA1. *J Neurosci* 22:2115–2124
 47. **Tanaka H, Yokota H, Jover T, Cappuccio I, Calderone A, Simionescu M, Bennett MV, Zukin RS** 2004 Ischemic preconditioning: neuronal survival in the face of caspase-3 activation. *J Neurosci* 24:2750–2759
 48. **Plahta WC, Clark DL, Colbourne F** 2004 17 β -Estradiol pretreatment reduces CA1 sector cell death and the spontaneous hyperthermia that follows forebrain ischemia in the gerbil. *Neuroscience* 129:187–193
 49. **Shughrue PJ, Merchenthaler I** 2003 Estrogen prevents the loss of CA1 hippocampal neurons in gerbils after ischemic injury. *Neuroscience* 116:851–861
 50. **Bondanelli M, Ambrosio MR, Onofri A, Bergonzoni A, Lavezzi S, Zatelli MC, Valle D, Basaglia N, Degli Uberti EC** 2006 Predictive value of circulating insulin-like growth factor I levels in ischemic stroke outcome. *J Clin Endocrinol Metab* 91:3928–3934
 51. **Harukuni I, Hurn PD, Crain BJ** 2001 Deleterious effect of β -estradiol in a rat model of transient forebrain ischemia. *Brain Res* 900:137–142
 52. **Espeland MA, Rapp SR, Shumaker SA, Brunner R, Manson JE, Sherwin BB, Hsia J, Margolis KL, Hogan PE, Wallace R, Dailey M, Freeman R, Hays J** 2004 Conjugated equine estrogens and global cognitive function in postmenopausal women: Women's Health Initiative Memory Study. *JAMA* 291:2959–2968
 53. **Rapp SR, Espeland MA, Shumaker SA, Henderson VW, Brunner RL, Manson JE, Gass ML, Stefanick ML, Lane DS, Hays J, Johnson KC, Coker LH, Dailey M, Bowen D** 2003 Effect of estrogen plus progestin on global cognitive function in postmenopausal women: the Women's Health Initiative Memory Study: a randomized controlled trial. *JAMA* 289:2663–2672
 54. **Sherwin BB** 2005 Estrogen and memory in women: how can we reconcile the findings? *Horm Behav* 47:371–375
 55. **Morrison JH, Brinton RD, Schmidt PJ, Gore AC** 2006 Estrogen, menopause, and the aging brain: how basic neuroscience can inform hormone therapy in women. *J Neurosci* 26:10332–10348
 56. **Dubal DB, Zhu H, Yu J, Rau SW, Shughrue PJ, Merchenthaler I, Kindy MS, Wise PM** 2001 Estrogen receptor α , not β , is a critical link in estradiol-mediated protection against brain injury. *Proc Natl Acad Sci USA* 98:1952–1957
 57. **Filardo EJ** 2002 Epidermal growth factor receptor (EGFR) transactivation by estrogen via the G-protein-coupled receptor, GPR30: a novel signaling pathway with potential significance for breast cancer. *J Steroid Biochem Mol Biol* 80:231–238
 58. **Thomas P, Pang Y, Filardo EJ, Dong J** 2005 Identity of an estrogen membrane receptor coupled to a G protein in human breast cancer cells. *Endocrinology* 146:624–632
 59. **Cardona-Gomez GP, Mendez P, Garcia-Segura LM** 2002 Synergistic interaction of estradiol and insulin-like growth factor-I in the activation of PI3K/Akt signaling in the adult rat hypothalamus. *Brain Res Mol Brain Res* 107: 80–88
 60. **Azcoitia I, Fernandez-Galaz C, Sierra A, Garcia-Segura LM** 1999 Gonadal hormones affect neuronal vulnerability to excitotoxin-induced degeneration. *J Neurocytol* 28:699–710
 61. **Yu X, Rajala RV, McGinnis JF, Li F, Anderson RE, Yan X, Li S, Elias RV, Knapp RR, Zhou X, Cao W** 2004 Involvement of insulin/phosphoinositide 3-kinase/Akt signal pathway in 17 β -estradiol-mediated neuroprotection. *J Biol Chem* 279:13086–13094
 62. **Cardona-Gomez GP, Trejo JL, Fernandez AM, Garcia-Segura LM** 2000 Estrogen receptors and insulin-like growth factor-I receptors mediate estrogen-dependent synaptic plasticity. *Neuroreport* 11:1735–1738
 63. **Quesada A, Etgen AM** 2002 Functional interactions between estrogen and insulin-like growth factor-I in the regulation of α 1B-adrenoceptors and female reproductive function. *J Neurosci* 22:2401–2408
 64. **Pietras RJ** 2003 Interactions between estrogen and growth factor receptors in human breast cancers and the tumor-associated vasculature. *Breast J* 9:361–373
 65. **Ronnekleiv OK, Kelly MJ** 2005 Diversity of ovarian steroid signaling in the hypothalamus. *Front Neuroendocrinol* 26:65–84
 66. **Kuroki Y, Fukushima K, Kanda Y, Mizuno K, Watanabe Y** 2001 Neuroprotection by estrogen via extracellular signal-regulated kinase against quinolinic acid-induced cell death in the rat hippocampus. *Eur J Neurosci* 13:472–476
 67. **Singer CA, Figueroa-Masot XA, Batchelor RH, Dorsa DM** 1999 The mitogen-activated protein kinase pathway mediates estrogen neuroprotection after glutamate toxicity in primary cortical neurons. *J Neurosci* 19:2455–2463
 68. **Fitzpatrick JL, Mize AL, Wade CB, Harris JA, Shapiro RA, Dorsa DM** 2002 Estrogen-mediated neuroprotection against β -amyloid toxicity requires expression of estrogen receptor α or β and activation of the MAPK pathway. *J Neurochem* 82:674–682
 69. **Mize AL, Shapiro RA, Dorsa DM** 2003 Estrogen receptor-mediated neuroprotection from oxidative stress requires activation of the mitogen-activated protein kinase pathway. *Endocrinology* 144:306–312
 70. **Alessandrini A, Namura S, Moskowitz MA, Bonventre JV** 1999 MEK1 protein kinase inhibition protects against damage resulting from focal cerebral ischemia. *Proc Natl Acad Sci USA* 96:12866–12869
 71. **Namura S, Ihara K, Takami S, Nagata I, Kikuchi H, Matsushita K, Moskowitz MA, Bonventre JV, Alessandrini A** 2001 Intravenous administration of MEK inhibitor U0126 affords brain protection against forebrain ischemia and focal cerebral ischemia. *Proc Natl Acad Sci USA* 98:11569–11574
 72. **Vartiainen N, Goldsteins G, Ketsa-Goldsteine V, Chan PH, Koistinaho J** 2003 Aspirin inhibits p44/42 mitogen-activated protein kinase and is protective against hypoxia/reoxygenation neuronal damage. *Stroke* 34:752–757
 73. **Cheung EC, Slack RS** 2004 Emerging role for ERK as a key regulator of neuronal apoptosis. *Sci STKE* 2004:E45
 74. **Wang RM, Zhang QG, Li CH, Zhang GY** 2005 Activation of extracellular signal-regulated kinase 5 may play a neuroprotective role in hippocampal CA3/DG region after cerebral ischemia. *J Neurosci Res* 80:391–399
 75. **Ospina JA, Brevig HN, Krause DN, Duckles SP** 2004 Estrogen suppresses IL-1 β -mediated induction of COX-2 pathway in rat cerebral blood vessels. *Am J Physiol Heart Circ Physiol* 286:H2010–H2019
 76. **Miller CA, Marshall JF** 2005 Molecular substrates for retrieval and reconsolidation of cocaine-associated contextual memory. *Neuron* 47:873–884
 77. **Guirland C, Suzuki S, Kojima M, Lu B, Zheng JQ** 2004 Lipid rafts mediate chemotropic guidance of nerve growth cones. *Neuron* 42:51–62
 78. **Impey S, Obrietan K, Wong ST, Poser S, Yano S, Wayman G, Deloulme JC, Chan G, Storm DR** 1998 Cross talk between ERK and PKA is required for Ca $^{2+}$ stimulation of CREB-dependent transcription and ERK nuclear translocation. *Neuron* 21:869–883
 79. **Bailey CH, Kaang BK, Chen M, Martin KC, Lim CS, Casadio A, Kandel ER** 1997 Mutation in the phosphorylation sites of MAP kinase blocks learning-related internalization of apCAM in Aplysia sensory neurons. *Neuron* 18:913–924
 80. **Martin KC, Michael D, Rose JC, Barad M, Casadio A, Zhu H, Kandel ER** 1997 MAP kinase translocates into the nucleus of the presynaptic cell and is required for long-term facilitation in Aplysia. *Neuron* 18:899–912
 81. **Zheng WH, Quirion R** 2006 Insulin-like growth factor-1 (IGF-1) induces the activation/phosphorylation of Akt kinase and cAMP response element-binding protein (CREB) by activating different signaling pathways in PC12 cells. *BMC Neurosci* 7:51
 82. **Goodenough S, Schleusner D, Pietrzik C, Skutella T, Behl C** 2005 Glycogen synthase kinase 3 β links neuroprotection by 17 β -estradiol to key Alzheimer processes. *Neuroscience* 132:581–589
 83. **Boulware MI, Weick JP, Becklund BR, Kuo SP, Groth RD, Mermelstein PG** 2005 Estradiol activates group I and II metabotropic glutamate receptor signaling, leading to opposing influences on cAMP response element-binding protein. *J Neurosci* 25:5066–5078
 84. **Dubal DB, Shughrue PJ, Wilson ME, Merchenthaler I, Wise PM** 1999 Es-

- tradiol modulates bcl-2 in cerebral ischemia: a potential role for estrogen receptors. *J Neurosci* 19:6385–6393
85. Alkayed NJ, Goto S, Sugo N, Joh HD, Klaus J, Crain BJ, Bernard O, Traystman RJ, Hurn PD 2001 Estrogen and Bcl-2: gene induction and effect of transgene in experimental stroke. *J Neurosci* 21:7543–7550
86. Ouyang YB, Tan Y, Comb M, Liu CL, Martone ME, Siesjo BK, Hu BR 1999 Survival- and death-promoting events after transient cerebral ischemia: phosphorylation of Akt, release of cytochrome C and activation of caspase-like proteases. *J Cereb Blood Flow Metab* 19:1126–1135
87. Sugawara T, Fujimura M, Morita-Fujimura Y, Kawase M, Chan PH 1999 Mitochondrial release of cytochrome c corresponds to the selective vulnerability of hippocampal CA1 neurons in rats after transient global cerebral ischemia. *J Neurosci* 19:RC39
88. Sugawara T, Noshita N, Lewen A, Gasche Y, Ferrand-Drake M, Fujimura M, Morita-Fujimura Y, Chan PH 2002 Overexpression of copper/zinc superoxide dismutase in transgenic rats protects vulnerable neurons against ischemic damage by blocking the mitochondrial pathway of caspase activation. *J Neurosci* 22:209–217
89. Bagetta G, Chiappetta O, Amantea D, Iannone M, Rotiroli D, Costa A, Nappi G, Corasaniti MT 2004 Estradiol reduces cytochrome c translocation and minimizes hippocampal damage caused by transient global ischemia in rat. *Neurosci Lett* 368:87–91

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